



Structural Changes in the Prostate of 3-Month-Old Rats with Chronic Alcoholism

1. Radjabov A. B.

Received 2nd Mar 2023,
Accepted 3rd Apr 2023,
Online 24th May 2023

¹ Bukhara State Medical Institute

Abstract: The article presents the results of a study on the histological relationship of glandular and non-glandular structures of the prostate gland of 3-month-old rats and its structural changes in chronic alcoholism.

In rats with chronic alcoholism, compared with the control, there is polymorphism of the terminal sections of the glands, an increase in the volume fraction of acini that do not contain a secret, and a decrease in the volume fraction of the glandular parenchyma in the structure of the organ. The experiment revealed acini with foci of epithelial stratification, vascular changes, mild and moderate lymphocytic infiltration, expansion of connective tissue layers, proliferation of fibrous connective tissue and compaction of bundles of collagen fibers.

Keywords: rat prostate, gland, morphometry, chronic alcoholism

INTRODUCTION

Among the priority problems in modern urology, with good reason, one can name the pathology of the prostate. One of the causes of prostate diseases lies in the structural and functional features of the tissues of this organ [3,4].

Chronic alcohol consumption is widespread in modern society. Alcohol is a dangerous toxic effect on most organs and systems. Alcohol abuse is the world's third risk factor for diseases and disability, is the cause of 60 types of diseases [5].

It should be noted that the morphology of the prostate in alcoholism is practically not studied. In this regard, the study of the morphological features of the organ in conditions of chronic alcoholism is of undoubted interest for theoretical and practical medicine.

MATERIALS AND METHODS

The study was performed on 24 outbred white male rats at the age of 3 months. 2 experimental groups were formed: 1st - control (n=12); 2nd – experimental group (n=12).

In the experimental group, for modeling chronic alcoholism, forced alcoholization of animals was used using a 40.0% ethanol solution. The solution was administered intragastrically using a metal probe 1

time per day at a total dose of 7 g/kg of body weight for 1 month before the study age. Control animals received intragastrically equal volumes of 0.9% NaCl solution. Rats were sacrificed by instantaneous decapitation under ether anesthesia, according to approved rules [3].

For histological examination, pieces of the prostate were fixed in 10% buffered formalin and embedded in paraffin according to the standard method. Histological sections obtained from paraffin blocks, 5-7 μm thick, were stained with hematoxylin and eosin for review purposes, collagen fibers were detected by van Gieson staining.

With a microscope magnification of 70 times (7x10), the sections were determined:

- the shape of the lumen of the glands, the number of terminal sections of the glands in the field of view, the volume fraction of acini with and without secretion (in %), the number of acini with desquamated epithelial cells in the field of view, in the intralobular stroma, the number of intraorgan vessels in the field of view was counted.

In the preparations, at a magnification of 280 times (7x40), using an eyepiece micrometer, the diameter of the lumen of the glands, the height of the epithelium, the inner diameter and wall thickness of intraorganic vessels were measured. In addition, the thickness of collagen fibers and their distribution in the tissues of the gland were determined.

In the field of view (7x40), the presence and severity of lymphocytic infiltration in the tissues of the gland was assessed. When distributing lymphocytes by severity (cell density), the classification of the North American Chronic Prostatitis Collaborative Research Network and the International Prostatitis Collaborative Network was used:

- 1) mild degree - single lymphocytic cells separated by distinct intermediate zones;
- 2) moderate degree - confluent fields of lymphocytic cells without tissue destruction and / or lymphoid nodular / follicular formation;
- 3) severe degree - confluent fields of lymphocytic cells with tissue destruction and / or lymphoid nodular / follicular formation.

To assess the severity (fibrosis) of the proliferation of connective tissue using an eyepiece micrometer with a magnification of the objective x40, eyepiece x7 in the field of view, the thickness of the stroma layers between the glands was measured.

The degree of compaction of the connective tissue was determined by the appropriate method (Gorbunova E.N., Davydova D.A., Krupin V.N., 2011) as follows: 1) mild form (increase in the thickness of stromal septa up to 2 times in 2-4 fields of view out of 10); 2) moderate form (the thickness of the stromal septa is increased up to 2 times in more than 4 fields of view or a sharp thickening - more than 3 times and is present in single (1-2) fields of view); 3) pronounced form (stromal septa are enlarged up to 3 times or more in 7-10 fields of view).

Conducted a study of the volume fractions of glandular and stromal elements (in%). To do this, using the morphometric grid G.G. Avtandilov (with the number of intersections 100) using an eyepiece x10, a lens x10 in each preparation of the prostate in 10 fields of view, the number of intersections falling on the stromal and glandular (including the lumen of the gland) elements was counted to determine their ratios.

RESULTS AND DISCUSSION

The study showed that in 3-month-old rats, the prostate has a normal structural plan, consists of numerous separate alveolar-tubular glands and muscular-elastic stroma in the form of loose fibrous connective tissue, bundles of smooth myocytes and vessels.

In survey microscopy, the terminal secretory sections or acini in most cases (75.0%) have a folded appearance, are represented by highly prismatic epithelium with high columnar and basal cells that rest on a clearly visible basement membrane. The height of the epithelial layer varies from 8.4 to 21.0 μm , on average $16.5 \pm 0.6 \mu\text{m}$. Acini have oval and rounded shapes. The diameter of the lumen of the glands ranges from 105.0 to 298.2 microns, on average - 198.7 ± 8.0 microns. The number of acini in the field of view ranges from 38 to 66, averaging 51.8 ± 1.5 . The volume fraction of acini with a secret is in the range of 70-100%, on average - 83.5 ± 1.6 . The proportion of acini without a secret is 8-30%, on average 16.5 ± 1.2 . The number of acini with desquamated epithelial cells in the lumen varies up to 4 per field of view, on average 1.7 ± 0.2 .

In the periglandular stroma, single lymphocytes are determined, separated by clear intervals. Their number in the field of view is in the range of 4-9, on average 5.7 ± 0.3 . The thickness of the stromal septa between the acini ranges from 8.4 to 25.2 μm , averaging $15.5 \pm 0.9 \mu\text{m}$.

The number of stromal vessels in the field of view is in the range of 3-8, averaging 5.0 ± 0.3 . The inner diameter of the venules is in the range from 16.8 to 19.4 microns, on average - 20.6 ± 0.67 microns. The thickness of their wall ranges from 4.2 to 8.4 microns, on average - 4.7 ± 0.21 microns. The diameter of the capillaries varies from 4.2 to 12.6 μm , on average $8.9 \pm 0.21 \mu\text{m}$. The wall thickness is in the range of 2.1-4.2 microns, on average - 4.0 ± 0.13 microns. The inner diameter of arterioles ranges from 8.2 to 16.4 μm , averaging $13.8 \pm 0.46 \mu\text{m}$. Their wall thickness varies from 4.2 to 8.4 μm , on average $7.5 \pm 0.21 \mu\text{m}$.

The volume fraction of glandular tissue is 62-80%, on average $71.0 \pm 1.0\%$. The proportion of stromal tissue ranges from 20-38%, averaging $29.0 \pm 1.0\%$.

Collagen fibers are located in the stroma of the prostate, forming a small-loop network, most of the fibers lie under the epithelium and surround the prostatic acini. The thickness of collagen fiber bundles varies from 4.2 to 12.6 μm , averaging $8.2 \pm 0.46 \mu\text{m}$.

It has been established that in 3-month-old rats of the experimental group, acini in 50% of cases have a folded appearance, are represented by low-prismatic epithelium, and high-prismatic epithelium is determined in some places. The height of the epithelium varies from 4.2 to 16.8 μm , on average, $9.1 \pm 0.6 \mu\text{m}$. In places in the epithelium, foci of epithelial stratification are defined, known as prostatic intraepithelial neoplasia (PIN). At the same time, the rows of layers in the epithelium are disturbed, cell polymorphism and the presence of large and multiple nucleoli in the nuclei are noted. Acini have a polygonal shape. The diameter of the lumen of the glands ranges from 25.2 to 126.0 microns, on average - 75.2 ± 0.4 microns. The number of acini in the field of view ranges from 54 to 96, averaging 74.2 ± 2.3 . The lumens of the acini are partially filled with a homogeneous secret. The volume fraction of acini with a secret is in the range of 15-46%, on average - 29.2 ± 1.7 . The proportion of acini without a secret is 54-85%, on average 70.8 ± 1.7 . In the lumen of individual acini, there are many fragments of desquamated, desquamated cells, their number varies from a few cells to small cell conglomerates. In the field of view, the number of acini with desquamated epithelium varies from 4 to 10, on average 7.5 ± 0.3 .

In the interacinar stroma, chaotically scattered single lymphocytes are determined in places. In most preparations, merging fields of lymphocytes are visualized; they envelop the end sections of the glands in the form of chains of round or oval shapes. In addition, accumulations of lymphocytes are found inside the lumen of the acini and around the vessels of the interlobular stroma, which infiltrate the walls of the vessels. At the same time, the integrity of the epithelial lining is not broken, tissue destruction and lymphoid nodules are not observed. The number of lymphocytes in the stroma (in the field of view) ranges from 15 to 40, on average 26.0 ± 1.4 . The thickness of the stromal septa ranges from 21.0 to 75.6 μm , averaging $40.7 \pm 2.94 \mu\text{m}$.

The number of stromal vessels in the field of view is in the range of 6-12, averaging 9.1 ± 0.3 . The inner diameter of the venules ranges from 16.8 to 37.8 μm , on average 25.2 ± 1.13 μm . The thickness of their wall ranges from 2.1 to 4.2 microns, on average - 3.57 ± 0.13 microns. The diameter of the capillaries varies from 8.4 to 16.8 μm , on average 12.6 ± 0.46 μm . The thickness of their wall is in the range of 2.1-4.2 microns, on average - 3.49 ± 0.13 microns. The inner diameter of arterioles ranges from 12.6 to 21.0 μm , averaging 17.5 ± 0.46 μm . Their wall thickness varies from 4.2 to 8.4 μm , on average 5.59 ± 0.21 μm .

Morphometry of the parenchymal-stromal ratio showed that the relative area of its parenchyma varies within 32-55%, averaging $40.6 \pm 1.24\%$. The proportion of stromal tissue ranges from 45-68%, averaging $59.4 \pm 1.24\%$.

In the experiment, numerous collagen fibers occupy all interepithelial regions of the stroma. They are found around the acini and ducts of the gland, where they densely braid the smooth myocytes of the stromal layer. In places, a coarse network of collagen fibers is formed in the interacinar stroma. The thickness of collagen fiber bundles varies from 4.2 to 8.4 μm , averaging 6.13 ± 0.21 μm .

CONCLUSION

Animals with chronic alcoholism have mild and moderate diffuse focal lymphocytic infiltration, vascular and alternative changes in the form of focal desquamation of varying severity. In some preparations, acini with foci of epithelial stratification are visualized, which is a precancerous condition.

In the experiment, a decrease in the size of acini, an increase in the volume fraction of acini that do not contain a secret, and a decrease in the volume fraction of the glandular parenchyma in the structure of the organ are observed.

The impact of alcohol leads to the growth of fibrous connective tissue and compaction of bundles of collagen fibers, which can be considered as a reaction of the body aimed at isolating the lesion from the surrounding tissues and systemic blood flow.

REFERENCES

1. Koptyaeva K.E., Muzhikyan A.A., Gushchin Ya.A., Belyaeva E.V., Makarova M.N., Makarov V.G. Method of autopsy and extraction of organs of laboratory animals // Message 1: rat. Laboratory animals for scientific research. - 2018. - No. 2. - C. 71-92. DOI: 10.29296/10.29296/2618723X-2018-02-08.
2. Sidorov P.I. The use of laboratory animals in a toxicological experiment: guidelines. Ed. Sidorova P.I. - Arkhangelsk. - 2002; 15 s.
3. Trotsenko B.V. Regional heterogeneity of the mesenchyme in the processes of prostate morphogenesis in human fetuses and rats / B.V. Trotsenko, I.A. Lugin // Morphology. 2009 V.3 No. 3. pp.126-130.
4. Linda M. Ernest Color Atlas of fetal and neonatal histology / M. Ernest Linda, Eduardo D. Ruchelli, Dale S. Huff // Springer. 2011. 399p.
5. Rocco A., Compare D., Angrisani D., Sanduzzi Zamparelli M., Nardone G. Alcoholic disease: liver and beyond / World J. Gastroenterol. 2014; 20(40): 14652-9.